

Iron HR L M227

0.1 - 10 mg/L Fe

Thioglycolate

Instrument specific information

The test can be performed on the following devices. In addition, the required cuvette and the absorption range of the photometer are indicated.

Instrument Type	Cuvette	λ	Measuring Range
, MD 600, MD 610, MD 640,	ø 24 mm	530 nm	0.1 - 10 mg/L Fe
XD 7000, XD 7500			

Material

Required material (partly optional):

Reagents	Packaging Unit	Part Number
KP962-Ammonium Persulphate Powder	Powder / 40 g	56P096240
Acidity / Alkalinity P Indicator PA1	30 mL	56L013530
Acidity / Alkalinity P Indicator PA1	65 mL	56L013565
Hardness Calcium Buffer CH2	65 mL	56L014465
Calcium Hardness Buffer CH2	5 x 65 mL mL	56L014472
Iron HR Reagent Set	1 pc.	56R023590

Application List

- · Cooling Water
- · Boiler Water
- Galvanization
- · Raw Water Treatment

Preparation

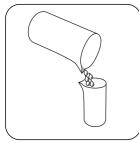
- If there are strong complexing agents in the sample, the response time must be extended until no further colour development is seen. However, very strong iron complexes are not included in the measurement. In this event, the complexing agent must be destroyed by means of oxidation with acid/persulphate and the sample also neutralised to pH 6–9.
- 2. For the measurement of total iron, both suspended and dissolved, the sample must be boiled with acid/persulphate. It must be neutralised back to pH 6–9 and refilled to the original volume with deionised water.





Digestion

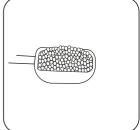
Total iron consists of suspended, soluble and complexed iron. The sample must be not filtered before measuring. To ensure homogenisation of the sample, deposed particles must be evenly distributed immediately prior to sampling by forcible shaking. A filtration of the sample is necessary for the determination of total soluble iron (including the complex iron compounds). The equipment required for the determination of total iron and reagents are not included in the standard delivery.



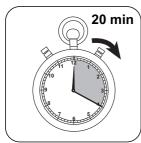
Fill a suitable digestion vessel with 50 mL homogenised sample.



Add 5 mL 1:1 Hydrochloric acid.



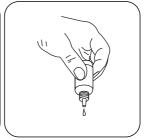
Add a measuring scoop KP 962 (Ammonium Persulphat Powder).



Boil the sample for **20 minutes**. A sample volume of about 25 mL should be retained; If necessary, fill with deionised water.



Allow the sample to cool to room temperature.

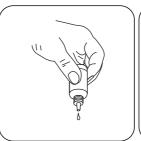


Hold cuvettes vertically and add equal drops by pressing slowly.





Add 1 drops Acidity / Add Hardness Calcium Alkalinity P Indicator PA1. Buffer CH2 drop by drop



Add Hardness Calcium Buffer CH2 drop by drop to the same sample until colouration turns from light pink to red. (Note: make sure to swirl the vial after adding each drop!)



Fill the sample with deionised water to 50 mL .



Determination of Iron, total HR with liquid reagent

Select the method on the device.

For testing of Iron, total HR with liquid reagent, carry out the described digestion.

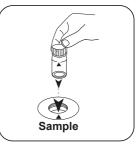
For this method, a ZERO measurement does not have to be carried out every time on the following devices: XD 7000, XD 7500

Total iron consists of suspended, soluble and complexed iron. The sample must be not filtered before measuring. To ensure homogenisation of the sample, deposed particles must be evenly distributed immediately prior to sampling by forcible shaking. A filtration of the sample is necessary for the determination of total soluble iron (including the complex iron compounds). The equipment required for the determination of total iron and reagents are not included in the standard delivery.



Fill 24 mm vial with 10 mL Close vial(s). deionised water.





Place sample vial in the sample chamber. Pay attention to the positioning.



Press the ZERO button.



Remove the vial from the sample chamber.



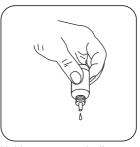
Empty vial.

For devices that require no ZERO measurement, start here.





Fill 24 mm vial with 10 mL prepared sample .



Hold cuvettes vertically and add equal drops by pressing slowly.



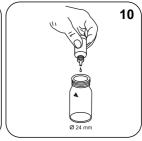
Add 10 drops Iron Reagent FE6.



Close vial(s).



Invert several times to mix the contents.



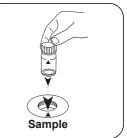
Add 10 drops Hardness Total Buffer TH2.



Close vial(s).



Invert several times to mix the contents.



Place **sample vial** in the sample chamber. Pay attention to the positioning.







Press the **TEST** (XD: **START**)button.

Wait for 15 minute(s) reaction time.

Once the reaction period is finished, the measurement takes place automatically.

The result in mg/L total Iron or when using a filtrated sample, in mg/l totale soluble Iron appears on the display.



Determination of Iron HR with Liquid Reagent

Select the method on the device.

For this method, a ZERO measurement does not have to be carried out every time on the following devices: XD 7000, XD 7500

For determination of dissolved iron the sample must be filtered prior to the test (pore size 0,45 µm). Otherwise, iron particles and suspended iron are measured.



Fill 24 mm vial with 10 mL Close vial(s).

Place sample vial in the sample chamber. Pay attention to the positioning.

sample.





Press the **ZERO** button.

Remove the vial from the sample chamber.

For devices that require no ZERO measurement, start here.



Hold cuvettes vertically and add equal drops by pressing slowly.



Add 10 drops Iron Reagent FE6.



Close vial(s).





Invert several times to mix the contents.



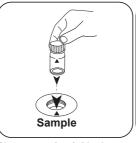
Add 10 drops Hardness Total Buffer TH2.



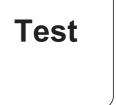
Close vial(s).



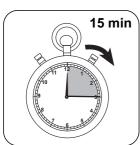
Invert several times to mix the contents.



Place **sample vial** in the sample chamber. Pay attention to the positioning.



Press the **TEST** (XD: **START**)button.



Wait for 15 minute(s) reaction time.

Once the reaction period is finished, the measurement takes place automatically.

The result in mg/L Iron appears on the display.



Chemical Method

Thioglycolate

Appendix

Calibration function for 3rd-party photometers

Conc. = a + b•Abs + c•Abs² + d•Abs³ + e•Abs⁴ + f•Abs⁵

	ø 24 mm	□ 10 mm
а	-1.53212 • 10 ⁻¹	-1.53212 • 10 ⁻¹
b	7.33471 • 10+0	1.57696 • 10+1
С		
d		
е		
f		

Bibliography

E. Lyons (1927), Thioglycolic Acid As A Colour Test For Iron, J. Am. Chem. Soc., 49 (8), p.1916-1920